Synthetic Approaches to the 2002 New Drugs

Jin Li* and Kevin K.-C. Liu*

Pfizer Global Research and Development, Pfizer Inc., Groton CT 06340, USA

Abstract: New drugs are introduced to the market every year and each individual drug represents a privileged structure for its biological target. In addition, these new chemical entities (NCEs) not only provide insights into molecular recognition, but also serve as drug-like leads for designing future new drugs. Therefore, it is important to be acquainted with these new structures as well as their syntheses. To these ends, this review covers the syntheses of 28 NCEs marketed in 2002.

Keywords: Synthesis, New Drug, New Chemical Entities, Medicine, Therapeutic Agents.

INTRODUCTION

Dozens of new drugs are registered and launched every year around the world. Although thousands of drugs have been marketed historically, the structure similarity among some drugs is obvious and even more so for drugs targeting in the same gene family. Furthermore, it has been demonstrated that molecules which share the same or similar chemical template can be further modified for different therapeutic indications against the similar gene family. Therefore, medicinal chemists, being aware of these new drug structures, can strike and adopt ideas for their own innovations. In addition, preparation of these drug molecules has been studied extensively to make it concise due to the cost of goods consideration and to ensure environmentfriendliness. Having such robust and reliable synthetic methods in hand to access these core structures will steer synthetic efforts more effectively toward the most promising compounds and help focus the optimization toward other challenging properties such as ADME.

In 2002 alone, 33 NCEs including biological drugs, and two diagnostic agents reached the market [1-5]. This review article will focus on the syntheses of the 28 new drugs marketed last year (Figure 1), but excludes new indications for known drugs, new combinations and new formulations. The syntheses of these new drugs were published sporadically in different journals and patents. It is our intention to compile the syntheses of new drugs yearly into an annual review for the readers' advantage. The synthetic routes cited here represent the most scalable methods according to the best of the authors' knowledge and appear in alphabetical order by generic name.

Adefovir Dipivoxil (HepseraTM)

Adefovir dipivoxil (1), discovered by Gilead, became the first nucleoside analogue to gain FDA approval for the treatment of chronic hepatitis B infection [6]. Adefovir works by blocking viral replication [6]. The synthesis [7,8] of adefovir dipivoxil (1) involves a four-step process [9,10] as depicted in Scheme 1. Adenine (29) was condensed with ethylene carbonate (30) in hot DMF to afford intermediate 9-

(2-hydroxyethyl)-adenine **31** in 83-95% yield. Alkylation of **31** was carried out using diethyl-*p*-toluenesulfonyloxymethanephosphonate (**32**) and sodium *t*-butoxide in DMF. Phosphonate ester **33** was then cleaved with bromotrimethylsilane to furnish **34** and esterification of the phosphoric acid to append the pivaloyloxymethyl group provided adefovir dipivoxil (**1**).

Amrubicin Hydrochloride (Calsed)

This drug is the first anthracycline anticancer antibiotic produced by purely synthetic methods. It was discovered by Sumitomo Pharmaceuticals, and is for the treatment of nonsmall cell lung cancer and small cell lung cancer [11]. Tetralone 35 was treated with ammonium carbonate and potassium cyanide (Strecker reaction) to give the corresponding aminonitrile intermediate, which was hydrolyzed under basic conditions to afford amino acid 36 in excellent yield [12]. The carboxylic acid in 36 was esterified with HCl in methanol to the corresponding methyl ester, which was treated with D-(-)-mandelic acid in toluene to give optically pure levorotatory ester 37 in 33% yield. Sodium methylsulfinylmethide treatment of 37 followed by reduction with zinc yielded amino ketone 38, which was acylated to give amido ketone 39 in 81 % yield from 37. Compound 39 was converted to tetracyclic amido ketone 40 in one step (90% yield) by heating with phthalic anhydride in the presence of AlCl₃-NaCl at 170°C. Ketone **40** was protected as its ketal 41 in order to provide for subsequent regiospecific bromination. Treatment of 41 with 1,3dibromo-5,5-dimethylhydantoin (DDH) under illumination in refluxing benzene formed oxazine 42 in 89% yield. Hydrolysis of the oxazine ring and deketalization were simultaneously affected by heating 42 with 3N sulfuric acid to give cis-amino alcohol 43 in 82% yield. Modified Arcamone conditions (AgOSO₂CF₃ in ether/tetramethylurea /DCM) were employed for the stereoselective glycosidation of 43 with 2-deoxy-3,4-di-O-acetyl-D-erythro-pentopyranosyl bromide (44) [13] to give the protected β-glycoside in 86% yield. Basic hydrolysis of the protected coupling product followed by HCl salt formation gave amrubicin hydrochloride (2) in 90% yield.

Aripiprazole (AbilifyTM)

This atypical antipsychotic agent was originally discovered by Otsuka and was co-developed and co-marketed

^{*}Address correspondence to these authors at the Pfizer, Groton, CT 06340, USA; Tel: 1-860-7153552; E-mail: jin_li@groton.pfizer.com; kevin_k_liu@groton.pfizer.com

(Fig. 1). contd.....

Pimecrolimus (21)

(Fig. 1). contd.....

Fig. (1). Structures of 28 new drugs marketed in 2002.

by Bristol-Myers Squibb. The compound is a partial agonist at dopamine D_2 and $5HT_{1a}$ and an antagonist at $5\text{-}HT_{2a}$ receptors [14]. It is indicated for the treatment of schizophrenia. Hydroxyl quinolinone **45** was alkylated with 1,4-dibromobutane in the presence of potassium carbonate in DMF to give **46** in 78% yield [15]. Bromide **46** was condensed with 1-(2,3-dichlorophenyl)piperazine [16] **(47)** in the presence of NaI and TEA to give aripiprazole **(3)** in 87% yield.

Dexmethylphenidate Hydrochloride (FocalinTM)

Dexmethylphenidate (4) is the more pharmacologically active *d-threo*-enantiomer of methylphenidate which was marketed for the treatment of attention deficit/hyperactivity disorder (ADHD) in 1954 [17]. In addition, it has been shown that there are significant metabolic differences between the two enantiomers. This drug was discovered by Cangene and is marketed by Novartis. To date, several methods have been disclosed in the literature for preparing

Scheme 1. Synthesis of adefovir dipivoxil.

Scheme 2. Synthesis of amrubicin hydrochloride.

the *d-threo*-enantiomer of methylphenidate, most involving with enzymatic resolution [18], or crystallization/recrystallization methods [19,20]. An asymmetric synthesis [21] route is depicted in Scheme 4. *R*-Pipecolic acid (48) was reacted with (Boc)₂O to afford *N*-Boc pipecolic acid 49. Treatment of 49 with *N*,*O*-dimethylhydroxylamine in DCM provided the Weinreb amide 50 in 93% yield. Reaction of amide 50 with phenyllithium at -23°C in Et₂O furnished enantiopure ketone 51 in 73% yield. Ketone 51 was converted to chiral aromatic alkene 52 using

methylenetriphenylphosphorium ylide in THF at rt. The transformation of olefin **52** to diastereomeric alcohols **53** and **54** was achieved using BH₃-THF complex in 89% overall yield. Diastereomerically pure alcohol **53** was subjected to PDC-mediated oxidation in DMF followed by treatment with excess ethereal diazomethane. The resulting *N*-Bocmethylphenidate was deprotected with 3N methanolic HCl to give dexmethylphenidate (**4**) as a white solid in 67% yield.

Scheme 3. Synthesis of aripiprazole.

OH
$$\frac{(Boc)_2O, TEA}{MeOH, rt, 97\%}$$
 OH $\frac{N, O\text{-dimethylhydroxylam ine}}{Boc}$ OH $\frac{N, O\text{-dimethylhydroxylam ine}}{BOP, TEA, DCM, π , 93% $\frac{N}{N}$ OH $\frac{N}{N}$ OH $\frac{N}{N}$ OH $\frac{N, O\text{-dimethylhydroxylam ine}}{BOP, TEA, DCM, π , 93% $\frac{N}{N}$ OH $\frac{N}{N}$$$

4

Scheme 4. Synthesis of dexmethylphenidate.

Dutasteride (AvodartTM)

53

This steroid 5α-reductase type 1 and 2 inhibitor was patented by GlaxoSmithKline. It is used for the treatment of symptomatic benign prostatic hyperplasia in men with an enlarged prostate to improve urinary symptoms, reduce the risk of acute urinary retention and BPH-related surgery [22]. Steroidal dutasteride (5) was synthesized from 3-oxo-4androstene-17β-carboxylic acid (55) [23]. Oxidation of 55 with potassium permanganate, sodium periodate and sodium carbonate in refluxing t-butyl alcohol and water gave secosteroid 56 which was cyclized with ammonium acetate in acetic acid to give 4-aza-steroid 57 in good yield. Stereoselective hydrogenation of 57 with H₂ over PtO₂ in hot acetic acid and in the presence of ammonium acetate yielded saturated azasteroid 58, which was dehydrogenated with DDQ in the presence of bis(trimethylsilyl)trifluoroacetamide (BSTFA) 59 in refluxing dioxane to give 60. Treatment of 60 with thionyl chloride gave the corresponding acyl chloride intermediate, which was then condensed with 2,5bis(trifluoromethyl)aniline (61) by means of DMAP in heated toluene to give dutasteride (5) in 57% yield from intermediate 60.

Ertapenem Sodium (InvanzTM)

Ertapenem sodium (6) was introduced in the U.S. and Europe by Merck & Co. as a once daily injectable carbapenum antibiotic drug. Ertapenem (6) is indicated for the treatment of moderate to severe infections in adults caused by susceptible strains of a range of Gram-positive and Gram-negative aerobic and anaerobic bacteria [24]. Following a conventional carbapenem synthetic strategy, ertapenem sodium (6) can be assembled from 4-nitrobenzylprotected β-methyl carbapenemenolphosphate 71 and 2aminocarbonylpyrrolidine-4-ylthio-containing side chain 70. Many efficient approaches to 71 have been reported in the literature [25], and this compound is now commercially available on a large scale [26]. The synthesis of 70 is outlined in Scheme 6 [27,28]. Protection of the amino group in trans-4-hydroxy-L-proline (62) with diisopropyl phosphite followed by NaClO oxidation gave N-DIPP protected hydroxyl proline 63 in 80% yield. The carboxyl group in 63 was activated via reaction with diphenylphosphinic chloride (DPPC) in the presence of diisopropylethylamine (DIPEA). This intermediate 64 was directly reacted with methanesulfonyl chloride in the

Scheme 5. Synthesis of dutasteride.

presence of pyridine to furnish mesylate 65. Mesylate 65 was then quenched with aqueous sodium sulfide yielding 66 instantaneously, which then slowly cyclized to 67. Aminolysis of 67 with m-aminobenzoic acid (68) and subsequent deprotection of the DIPP group with concentrated HCl provided 70 in 90-95% yield in a one-pot process. The coupling reaction between 70 and 71 followed by deprotection of PNB group was completed in one reaction vessel to furnish ertapenem sodium (6) (yield was not disclosed) [28].

Escitalopram Oxalate (Cipralex®)

Escitalopram (7) is a selective serotonin reuptake inhibitor (SSRI) and was launched first in Switzerland. It is the more active S-enantiomer of citalogram which is a wellknown antidepressant drug that has been on the market for some years [29]. It is for the treatment of major depressive episodes and panic disorder with or without agoraphobia. The synthesis of escitalopram was carried out in several different routes [30-33]. 5-Cyanophthalide (72) was treated with Grignard reagent 73 at 0°C to provide intermediate 75 which was reacted in situ with another Grignard reagent 76 to afford the diol in a one-pot process. Racemic diol 77 was resolved using (+)-p-toluoyltartaric acid to afford desired S isomer 78 in 55% yield. The ring closure reaction was carried out at 0°C using methanesulfonyl chloride in toluene to furnish escitalopram (7) in 60% yield.

Etoricoxib (ArcoxiaTM)

Merck & Co.'s etoricoxib (8) was launched for the first time in the U.K. last May as a new COX-2 inhibitor.

Etoricoxib (8) is indicated for the symptomatic relief of osteoarthritis and rheumatoid arthritis, treatment of acute gouty arthritis, relief of chronic musculoskeletal pain including low back pain, relief of acute pain associated with dental surgery and treatment of primary dysmenorrhea [34]. The synthesis of etoricoxib (8) was explored extensively by the Merck process research group [35]. Key intermediate 85 was synthesized through at least three different routes. In the Horner-Wittig approach, 6-methyl methylnicotinate (79) was converted into Weinreb amide 80 in 95% yield. Amide 80 was then converted to aldehyde 81 via a DIBAL-H mediated reduction. Subsequent treatment of a solution of aldehyde 81 in isopropyl acetate with aniline and diphenyl phosphite provided N,P-acetal 82 in 87% yield. The Horner-Wittig reaction of N, P-acetal 82 with 4-methanesulfonylbenzaldehyde (83) furnished enamine 84, which was hydrolyzed to ketosulfone 85. A Grignard approach was also developed in the preparation of ketosulfone 85. Addition of Grignard reagent 86 to Weinreb amide 80 in toluene/THF provided ketosulfide 85 in 80% yield. Tungstate-catalyzed oxidation of ketosulfide 87 using hydrogen peroxide provided ketosulfone 85 in 89% yield by simple filtration. Ketosulfone **85** was prepared through Claisen condensation protocol as well. Thus, reaction of 4-methanesulfonyl phenyl acetic acid (88) with methyl nicotinate 79 under Ivanoff condition, i.e., the magnesium dianion in THF, resulted 58% yield of ketosulfone 85. Treatment of ketosulfone 85 with a three-carbon electrophile, 2-chloro-N, Ndimethylaminotrimethinium hexafluorophos-phate (89) in the presence of potassium *t*-butoxide at ambient temperature resulted adduct 90. Inverse quench of adduct 90 into a mixture of HOAc /TFA led to the putative intermediate 91. Ring closure of the pyridine ring occurred upon heating at

$$H_2N$$
 H_2N
 H_2N
 H_3
 H_4
 H_5
 H_5
 H_5
 H_5
 H_6
 H_7
 H

one pot process, 90-95% from 67 to 70

Scheme 6. Synthesis of ertapenem sodium.

reflux in the presence of an excess of aqueous ammonium hydroxide to give desired etoricoxib (8) in 97% yield in a one-pot process from 85.

Ezetimibe (Zetia)

Ezetimibe (9) was approved as the first hypolipidemic drug to act by blocking the absorption of dietary cholesterol. This drug was discovered by Schering-Plough and is codeveloped and co-marketed by Merck and Schering-Plough for the treatment of hypercholesterolemia and also two less common forms of hyperlipidemia: homozygous familial hypercholesterolemia and homozygous sitosterolemia [36]. The synthesis of ezetimibe (9) begins with the one-step diastereoselective and practical synthesis [37] of the trans βlactam from commercially available (S)-3-hydroxy- γ -lactone (92). Lactam 95 was obtained by generation of a dianion of lactone 92 with LDA in THF followed by addition of the imine and N, N'-dimethylpropyleneurea (DMPU) to give predominately adduct 93 (93:94 = 79:21). However, intermediate 93 and 94 did not cyclize to their respective lactams due to formation of stable lithium aggregates.

Addition of lithium chloride/DMF was employed to cyclize the intermediates into trans-lactam 95 as the major product (trans: cis = 95:5) in a one-pot process from 92 in 64% yield. The 95:5 ratio of compound 95 was oxidatively cleaved with NaIO₄ to give aldehyde **96**. Mukaiyama aldol condensation was adopted to elaborate the 4-fluorophenylpropyl side chain to give alcohol 98. Without isolation, the reaction mixture was subjected to dehydration using p-TSA to give enone 99 in 75% yield from compound 96. Reduction of the double bond in 99 with Wilkinson's catalyst yielded ketone 100, which was subjected to the highly enantioselective CBS reduction to give alcohol 101 with a 98:2 selectivity of S:R at the benzylic position. Catalytic hydrogenation of compound 101 gave ezetimibe (9) in 79% yield. Alternatively, a palladium-catalyzed double reduction in EtOAc/MeOH of both the double bond and the benzyl protecting group in enone 99 produced free phenol 107 in 90% yield. A three-step one-pot procedure was subsequently developed to transform 107 into ezetimibe (9) in 79% yield. That is, free phenol 107 was protected in situ as its TMS ether using BSU followed by a highly selective CBS reduction of the ketone group to give the

Scheme 7. Synthesis of escitalopram.

desired alcohol in 97% ee. The TMS group was removed during acidic workup to give ezetimibe (9). A more convergent approach to this drug was also developed by preparing the (S)-hydroxy side chain before the ring construction [38]. Therefore, p-fluorobenzovlbutyric acid (102) was reacted with pivaloyl chloride and the acid chloride thus obtained was acylated with chiral auxiliary 103 to give the corresponding amide. The ketone group in the amide was reduced with (R)-MeCBS/BH₃-THF (104) in the presence of p-TSA to give desired alcohol 105 in high yield (99%) and stereoselectivity (96 % d.e.) [39]. Chiral alcohol 105 was then mixed with the imine in the presence of TMSCl and DIPEA to protect the alcohols as TMS ethers. In the same pot, TiCl₄ was added to catalyze the condensation reaction and gave compound 106 in 65% yield. Compound 106 was reacted with TBAF and a fluoridecatalyzed cyclization took place to give the corresponding lactam. Finally, the TMS protecting group was removed under acidic conditions to give ezetimibe (9) in 91% yield over two steps.

Fondaparinux Sodium (ArixtraTM)

Fondaparinux sodium (Arixtra; formerly fondaparin sodium, 10) is a synthetic pentasaccharide heparinoid Factor Xa antagonist and thrombokinase inhibitor launched extensively by Sanofi-Synthélabo (formerly Sanofi) and Organon as a treatment and prophylaxis for deep vein thrombosis (DVT) and symptomatic pulmonary embolism following hip or knee surgery. It is also being developed as a potential treatment for coronary artery diseases [40]. Fondaparinux has a complex structure. Starting from Dglucose, D-cellobiose, and D-glucosamine, the production process for the synthesis of the pentasaccharide involves about 55 steps. The synthesis was accomplished by preparing a fully-protected pentasaccharide, and then converting it into the final product. The choice of protecting groups was dictated by two factors: the need to introduce sulfate substituents (O- as well as N-linked), carboxylate groups and hydroxyl groups, in the proper positions on the target molecule, and the constraints of current methods for oligosaccharide synthesis, particularly the use of 2-azido glucose derivatives to achieve stereoselective introduction of α-D-linked glucosamine units. All the monosaccharide synthons were obtained from glucose or from glucosamine [41,42], and the synthesis [42-44] is outlined in Scheme 10. Trisaccharide 108 and disaccharide 109 are the two key building blocks in the synthesis. Coupling 108 and 109 was carried out at -20°C in DCE. Fully protected pentasaccharide 110 was then converted into the target compound 10 using traditional methods: saponification, O-sulfation, cleavage of benzyl ethers with simultaneous reduction of azido into

82

85
$$\frac{\text{Cl}}{\text{NMe}_2}$$
 $\frac{\text{SO}_2\text{Me}}{\text{N}^+}$ $\frac{\text{Cl}}{\text{N}^+}$ $\frac{\text{N}^+}{\text{N}^+}$ $\frac{\text{Cl}}{\text{N}^+}$ $\frac{\text{N}^+}{\text{N}^+}$ $\frac{\text{Cl}}{\text{N}^+}$ $\frac{\text{N}^+}{\text{N}^+}$ $\frac{\text{N}^+}{\text{N}^+}$

Scheme 8. Synthesis of etoricoxib.

amino functions and finally *N*-sulfation. Preparation of trisaccharide building block **108** started from 1,6-anhydrocellobiose (**111**). Selective protection at 4',6' position was achieved through benzylidenation to provide crude **112** which was converted into epoxide **113** by treatment with sodium methoxide and benzylation. Compound **113** was

isolated after filtration on silica gel and crystallization (m.p. 184-5°C). *Trans*-diaxial opening of the epoxide yielded the 2-azido derivative (66%) which was acetylated to give **114** (99%). The benzylidene was cleaved (92%) and the diol was then converted into **115** by successive tritylation, levulinoylation, detritylation, oxidation, methylation and

hydrazinolysis (60% over the 6 steps). Imidate 116 was prepared in the usual way from its hydroxyl precursor and coupled with 115 to give O-linked trisaccharide 117 in 78% yield. Compound 117 was acetolysed (91%), the anomeric acetate was cleaved by benzylamine in ether (100%) and imidate 108 was obtained by reaction with potassium carbonate and trichloroacetonitrile at room temperature (α ,

 β – mixture with α as the predominant isomer, 76%). The preparation of the other building block **109** is described as following. Selective 6-acetylation of **118** by *N*-acetylimidazole in DCE gave **119** in 60% yield. Treatment of **119** with **120** using DCE/pyridinium perchlorate and followed dechloroacetylation using hydrazinedithiocarbonate afforded the crystalline disaccharide **109** [43].

Scheme 9. Synthesis of ezetimibe.

Frovatriptan Succinate (FrovaTM)

The serotonin 5-HT_{1D} receptor agonist frovatriptan succinate (11) was launched last year in the U.S. for the acute treatment of migraine attacks. This drug was discovered at Vernalis and is marketed by UCB Pharm and Elan. Frovatriptan treats migraine by constricting blood vessels in the brain [45]. The synthesis of frovatriptan (11) appeared in a patent in multi-kilo scale [46]. Cyclohexanedione monoketal (121) was converted to amine 122 by reductive amination. The Fischer indolization of amine 122 with hydrazine 123 furnished indole nitrile 124 in 72% yield. The desired R isomer of the indole nitrile was obtained via a chiral salt formation/recrystallization process using chiral lactam 125 and isolated as a L-pyroglutamic acid salt 126. Hydrolysis of the nitrile functional group in 126 provided carboxamido indole 127, which was converted to succinate 11 in situ.

Fulvestrant (Faslodex®)

Fulvestrant (12) was launched for the first time in the U.S. for the treatment of hormone receptor-positive metastatic breast cancer in postmenopausal women with disease progression following antiestrogen therapy. As an

estrogen antagonist with no known agonist effects, it is the only compound in its class to be proven effective after tamoxifen failure [47]. It is administered as a once a month i. m. injection. Several routes for the synthesis of fulvestrant (12) were published [48,49]. One of the best routes [50] is depicted in Scheme 12. The conjugate addition of Grignard reagent derived from bromide 130 with dienone 129 gave adduct 131 as a mixture of 7α - and 7β -isomers in a ratio of 2.5:1 in 90-95% yield. Aromatization of the A-ring with copper bromide/lithium bromide in acetic acid followed by hydrolysis of the ester group provided diol 132 in 80-85% yield. Oxidation of the side chain from sulfite to sulfone followed by crystallization provided fulvestrant (12) in 30% overall yield from dienone 129.

Gefitinib (Iressa)

Gefitinib (13) is the first drug in a new class of anticancer agents known as epidermal growth factor receptor (EGFR) inhibitors. It was discovered by AstraZeneca and is for the treatment of inoperable or recurrent non-small cell lung cancer [51]. A mixture of 4,5-dimethoxyanthranilic acid (133) and formamide was heated to generate the cyclized quinazoline 134 [52]. The quinazoline was selectively mono-

Scheme 10. Synthesis of fondaparinux sodium.

Scheme 11. Synthesis of frovatriptan.

OAc

$$A = H$$
 $A = H$
 $A = H$

Scheme 12. Synthesis of fulvestrant.

demethylated with methionine in refluxing methanesulfonic acid to afford 135 in 47% yield [53]. Compound 135 was acylated to give acetate 136, which was treated with refluxing thionyl chloride to yield chloropyrimidine 137. Chloride 137 was condensed with 3-chloro-4-fluoroaniline (138) in refluxing IPA to yield anilinoquinazoline 139 in

56% yield from 136. The acetate protecting group in compound 139 was hydrolyzed with ammonium hydroxide in methanol, and the free phenol was alkylated with 3-(4-morpholinyl)propyl chloride (140) to give gefitinib (13) in 55% yield.

13

Scheme 13. Synthesis of gefitinib.

Landiolol Hydrochloride (Onoact®)

Landiolol hydrochloride (14) was launched in Japan by Ono for the treatment of intraoperative tachyarrhythmia. It improves tachyarrhythmia by selectively blocking β_1 receptors located mainly in the heart and by inhibiting the action of catecholamine [54]. The synthesis of landiolol appeared in an earlier patent in 1990 [55]. Esterification of 3-(4-hydroxyphenyl)propionic acid (141) with 2,2-dimethyl-

1,3-dioxolan-4-ylmethyl chloride (142) in DMSO gave desired ester 143 in 57% yield. Treatment of phenol 143 with bromo epoxide 144 in the present of K_2CO_3 afforded ether 145 in 76% yield. Epoxide 145 was then reacted with free amine 146 *via* a neucleophilic ring opening process to provide landiolol (14).

Micafungin Sodium (Funguarg®)

Scheme 14. Synthesis of landiolol.

Scheme 15. Synthesis of micafungin sodium.

The semi-synthetic echinocandin antifungal agent, micafungin sodium (15), is a 1,3-β-glucan synthase inhibitor discovered by Fujisawa. It is for the treatment and prevention of infections caused by *Aspergillus* and *Candida* such as *fungemia*, respiratory mycosis and gastrointestinal mycosis [56]. The key intermediate for the side chain of micafungin (15) was prepared by regioselective 1,3-dipolar cycloaddition reaction of 4-methoxycarbonylbenzhydroxamic acid chloride (147) and 4-pentyloxyphenylacetylene (148) with TEA in THF [57]. Basic hydrolysis of thus obtained ester 149, followed by condensation with 1-hydroxybenzotriazole (HOBT) gave the corresponding

activated ester 150 in 95% yield. The cyclic peptide core 151, obtained by acylase-catalyzed hydrolysis of the natural product FR901379, was acylated with 150 to give micafungin (15) in 53% yield.

Neridronate (Nerixia®)

This bisphosphonate compound was developed, and is marketed, by Abiogen Pharma. This drug is the first treatment ever for osteogenesis imperfecta [58]. 6-Aminohexanoic acid (152) was reacted with phosphorus trichloride and phosphorous acid at 85°C, and then water

Scheme 16. Synthesis of neridronate.

Scheme 17. Synthesis of nitisinone.

was added to generate free diphosphonic acid **16** in 78% overall yield [59].

Nitisinone (Orfadin®)

This reversible inhibitor of 4-hydroxyphenylpyruvate dioxygenase was discovered by Swedish Orphan and is comarketed by Apoteket AB and Rare Disease Therapeutics. It is used as an adjunct to dietary restriction of tyrosine and phenylalanine in the treatment of hereditary tyrosinemia type 1 (HT-1) disease [60]. Nitisinone (17) was synthesized in one step by reacting 2-nitro-4-trifluoromethylbenzoyl chloride (153) with cyclohexane-1, 3-dione (154) in the presence of TEA and trimethylsilylcyanide or 2-cyano-2-(trimethylsilyloxy)propane [61].

Olmesartan Medoxomil (Benicar TM)

This Angiotensin II antagonist was discovered by Sankyo and licensed to Forest for the treatment, alone or in combination with other antihypertensive agents, of high blood pressure [62]. The imidazole ring of olmesartan (18) was constructed with diaminomaleonitrile 155 and trimethylorthobutyrate (156) in CH₃CN then xylene to give 157 in 96% yield [63]. Acid hydrolysis of 157 in 6N HCl gave the dicarboxylic acid intermediate. After esterification of the diacid in ethanol in the presence of HCl, diester 158 was treated with MeMgCl to give 4-(1-hydroxyalkyl) imidazole 159 in 95% yield. Alkylation of 159 with biphenyl bromide 160 in the presence of potassium *t*-butoxide afforded 161 in 80% yield. Ester 161 was then hydrolyzed to free carboxylic acid 162 under basic

Scheme 18. Synthesis of olmesartan medoxomil.

Scheme 19. Synthesis of paracoxib sodium.

conditions, and 162 was treated with chloride 163 in the presence of K_2CO_3 to give ester 164 in 88% yield from 161.

Lastly, the trityl group was removed with 25% aqueous acetic acid to give olmesartan (18) in 81% yield.

Pazufloxacin mesi late

Scheme 20. Synthesis of pazufloxacin mesilate.

Parecoxib Sodium (Dynastat®)

Parecoxib sodium (19) is a cyclooxygenase 2 (COX-2) inhibitor and was introduced by Pharmacia (now Pfizer) as an injectable formulation for short-term treatment of postoperative pain [64]. Parecoxib is a water-soluble prodrug of valdecoxib (27) that undergoes biotransformation in vivo to release valdecoxib (27). The synthesis (Scheme 19) of parecoxib sodium (19) started from commercially available deoxybenzoin (165). Deoxybenzoin (165) was treated with hydroxylamine in EtOH/H₂O (3:1) to give deoxybenzoin oxime 166 in 95% yield. Deprotonation of oxime 166 with two equivalents of *n*-hexyllithium followed by condensation with ethyl acetate afforded isoxazoline 167 in 59% yield. Treatment of isoxazoline 167 with chlorosulfonic acid followed by reaction of the incipient sulfonyl chloride with aqueous ammonia furnished valdecoxib (27). Acylation of isoxazole sulfonamide 27 with propionic anhydride afforded parecoxib, which was converted to its sodium salt by titration with aqueous sodium hydroxide (64%) [65,66].

Pazufloxacin Mesilate (Pazucross, Pasil)

This fluoroquinolone was co-developed by Toyama and Mitsubishi Pharm and was launched for the intravenous therapy of respiratory, urinary, surgical, gynecological and systemic infections [67]. The drug is elegantly synthesized

from commercially available 2,3,4,5-tetrafluorobenzoic acid (168) by an 11-step process with an overall yield 48% [68]. Starting material 168 was first treated with ethyl bromide and then with t-butyl cyanoacetate in the presence of potassium carbonate in DMSO in one flask to give acylated cyanoacetate 169. Intermediate 169 thus obtained without purification was refluxed in toluene with p-TSA to yield 4cyanomethylbenzoate 170 in 90% yield from 168. Cyclopropanation at the benzylic position of 170 was performed by α,α -dialkylation with two equiv. of 1,2dibromoethane under phase-transfer conditions to give cyanocyclopropyl compound 171. Cyano compound 171 was subjected to hydration with alkaline H₂O₂ to afford carboxamide 172 in 81% yield from 170. Subsequently, carboxamide 172 was treated with NaOCl for Hofmann rearrangement to give primary amine 173, which was protected as its N-acetyl derivative 174 for the next reaction. Treatment of 174 with imidazole in the presence of thionyl chloride and TEA generated an imidazolide intermediate, which was converted to β-keto ester 175 by reacting with potassium ethyl malonate and MgCl₂. Enamine 176 was obtained without purification by successive treatment of 175 with DMF-dimethylacetal and (S)-(+)-2-aminopropanol. Crude 176 was heated in DMSO in the presence of potassium carbonate to efficiently give tricycle product 177 in 80% yield from 174. Finally, the ethyl ester and

Scheme 21. Synthesis of pimecrolimus.

acetamide in 177 were hydrolyzed under basic and acidic conditions, respectively, to give the free amine. Pazufloxacin mesilate (20) was obtained in 94% yield by treatment of its corresponding free amine with methanesulfonic acid in ethanol.

Pimecrolimus (Elidel®)

Pimercrolimus (21) is the first non-steroid agent for the treatment of mild to moderate atopic dermatitis lunched by Novartis. It selectively blocks the production and release of cytokines from T-cells. These cytokines cause inflammation, redness and itching associated with eczema. Long-term therapy with pimecrolimus (21) was more effective than conventional treatment in reducing the incidence of disease flares and the use of corticosteroids. This drug is also safe and effective in pediatric patients and is approved for use in children as young as two years [69]. The syntheses of pimecrolimus (21) appeared in several patent applications [70-73]. Starting material 178 was prepared by either fermentation [74] or modification of a previously described synthetic method in the literature [75]. Treatment of

macrolide 178 with triisopropylsilyl trifluoromethane-sulfonate (TIPS-triflate) in the presence of lutidine in DCM at 0°C afforded di-protected compound 179 in 94% yield. Selective deprotection of the TIPS group at position 32 using *p*-TSA in MeOH at rt gave mono-protected macrolide 180 in 88% yield. Reaction of the hydroxyl group at position 32 with *o*-nitrobenzenesulfonyl chloride (181) in the presence of DMAP and DIPEA in DCM provided 182 in 78% yield with 20% recovered starting material 180. Displacement of the sulfate with chloride using LiCl in DMF furnished the chlorinated compound, which was treated with aqueous HF to remove the TIPS group to provide pimecrolimus (21).

Prulifloxacin (Sword)

This fluoroquinolone antibacterial prodrug was originally discovered by Nippon Shinyaku and subsequently codeveloped and co-marketed by Meiji Seika. The drug is used in the treatment of systemic bacterial infections including acute upper respiratory tract infection, bacterial pneumonia, cholecystitis, prostatitis, internal genital infections, bacterial

Prulifloxacin

Scheme 22. Synthesis of prulifloxacin.

enteritis, otitis media and sinusitis [76]. The synthesis of prulifloxacin (22) [77] started with the treatment of 3,4difluoroaniline (183) with carbon disulfide in the presence of TEA to give the triethylammonium dithiocarbamate, which by reaction with ethyl chloroformate and TEA in chloroform, was converted into isothiocyanate 184 in 74% yield. Reaction of 184 with diethyl malonate in the presence of KOH in dioxane yielded methylenemalonate 185 potassium salt, which was ethylated with ethyl sulfate in ethanol to give compound 186 in excellent yield. 6,7-Difluoroquinoline 187 was obtained with the highest yield and regioselectivity when precursor 186 was heated in refluxing xylene [78]. To suppress the side reaction in the subsequent chlorination, quinoline 187 was acylated to give 188 with acetyl chloride in chloroform. Chlorination of 188 with sulfuryl chloride gave compound 189 in 79% yield. Compound 189 was treated with sodium acetate in THF to afford cyclized compound 190, which was condensed with piperazine in DMF to give compound **191**. The hydrolysis of ester **191** with KOH in hot *t*-butanol gave free acid **192**, which was finally condensed with 4-(bromomethyl)-5-methyl-1, 3-dioxol-2-one (**163**) by treatment of potassium bicarbonate in DMF to give prulifloxacin (**22**).

Rosuvastatin Calcium (Crestor®)

The HMG-CoA reductase inhibitor, known as Crestor® (23), was originally discovered by Shionogi and subsequently co-developed and co-marketed by AstraZeneca. The drug is for the treatment of patients with primary hypercholesterolemia (type IIa including heterozygous familial hypercholesterolemia) or mixed dyslipidemia (type IIb) as an adjunct to diet when response to exercise and diet is inadequate. Crestor (23) is also used in patients with homozygous familial hypercholesterolemia either alone or as an adjunct to diet and other lipid-lowering treatments [79].

Scheme 23. Synthesis of rosuvastatin calcium sodium.

Scheme 24. Synthesis of sivelestat sodium hydrate.

The synthesis of optically pure rosuvastatin (23) begins from the Knoevenagel reaction of p-fluorobenzaldehyde (193) with ethyl isobutylacetate (194) to give unsaturated ketoester 195 [80]. Compound 195 was condensed with (S)methylisothiourea and then aromatized in situ using DDQ in methylene chloride to give pyrimidine 196 in 50% yield. Pyrimidine sulfide **196** was then oxided by *m*-CPBA to give sulfone 197 in 96% yield. Sulfone 197 was reacted with methylamine in methanol followed by treatment with methanesulfonyl chloride to give the methanesulfonylamino pyrimidine 198 in 58% yield. Reduction of ester 198 with DIBAL-H followed by TPAP oxidation afforded aldehyde 199 in 58% yield. Aldehyde 199 was subjected to Wittig reaction with optically pure ylide, (3*R*)-3-(*t*-butyldimethylsilyloxy)-5-oxo-6-triphenylphosphoranylidenehexanoate (200) [81], to give heptenoate compound 201 in 71% yield. Compound 201 was deprotected with HF in acetonitrile, and stereoselective chelation-controlled reduction with Et_2BOMe and $NaBH_4$ in THF-MeOH mixed solvent gave methyl (3R, 5S, 6E)-dihydroxyheptenoate **202** in 85% yield. Diol **202** was hydrolyzed with aqueous NaOH to afford the corresponding sodium salt. Rosuvastatin calcium salt (**23**) was obtained as white powder from the sodium salt on treatment with aqueous $CaCl_2$.

Sivelestat Sodium Hydrate (Elaspol®)

A neutrophil elastase inhibitor, introduced by Ono Pharmaceutics as an injectable formulation, is for the treatment of acute lung injury accompanying systemic inflammatory response syndrome [82]. The synthesis [83] of sivelestat (24) started with the amide formation between 2-nitrobenzoyl chloride (203) and glycine benzyl ester *p*-tolene sulfonic acid salt (204) in the presence of TEA to give amide 205 in 90% yield. Amide 205 was then reduced with iron

Scheme 25. Synthesis of tiotropium sodium.

Scheme 26. Synthesis of treprostinil sodium.

power under acidic conditions to give corresponding amine **206** in 81% yield. Alternatively, the mixture of activated Raney nickel, nitro compound **205**, acetic acid and 1,3-dimethyl-2-imidazolinone (DMI) under 25 atmospheric pressure of hydrogen at 40°C in an autoclave can give the same free amine **206** in 88% yield. Free amine **206** was

treated with p-pivaloyloxybenzenesulfonyl chloride [84] (207) in pyridine to yield sulfonamide 208 in 87% yield. Benzyl ester 208 was converted to its free carboxylic acid under hydrogenation, and the carboxylic acid was subsequently basified to give sivelestat sodium (24).

Tiotropium Bromide (Spiriva®)

Boehringer Ingelheim's once-daily inhaled chronic obstructive pulmonary disease (COPD) therapy tiotropium bromide (25) was launched for the first time in the Netherlands and Philippines in 2002. Tiotropium (25), which acts through prolonged M3 receptor blockade, is approved as a bronchodilator for the maintenance treatment of COPD [85]. At least two synthetic paths have been disclosed in the patent and literature [86-88]. The synthesis of tiotropium is depicted in Scheme 25. Tropenol hydrochloride 209 was first neutralized with ammonia in toluene and then the free base was reacted with methyl di-(2thienyl)glycolate (210) in the presence of sodium hydride to furnish desired tropenol ester 211 in 83% yield. The vanadium-catalyzed oxidation of tropenol ester 211 using hydrogen peroxide-urea complex gave epoxide 212, which was converted into its quaternary salt 25 with methyl bromide. The last two steps were carried out in a one-pot process in 88% yield.

Treprostinil Sodium (RemodulinTM)

The prostacyclin analog, treprostinil sodium (26), was launched in the U.S. in June 2002 for the treatment of pulmonary hypertension. Developed and marketed by United Therapeutics, treprostinil is specifically approved for the treatment of pulmonary arterial hypertension in patients with NYHA class II-IV symptoms, to reduce symptoms associated with exercise [89]. The synthesis of treprostinil [90,91] starts from commercially available 3-methoxybenzyl alcohol (213). The hydroxyl group in 213 was protected as a t-butyldimethylsilyl ether via reaction with TBDMS chloride in DCM at rt. A regiospecific introduction of the allylic chain and deprotection of the silyl group in situ provided alcohol 216 in 36% yield in a three-step sequence. Swern oxidation of alcohol 216 using oxalyl chloride/DMSO furnished aldehyde 217 in 86% yield. Acetylene 218 was first treated with magnesium ethyl bromide and then reacted with aldehyde 217 to provide adduct 219 in 52% yield. The alcohol functional group in

Scheme 27. Synthesis of voriconazole.

219 was then transformed into a carbonyl group in 220 via a PCC-mediated oxidation. Ketone 220 was then reduced again using chiral boron reagent to give the chiral alcohol which was protected with TBDMS chloride in situ (221). Optically pure intermediate 221 underwent cobalt-mediated Pauson-Khand reaction to furnish tricyclic compound 222 in excellent yield. Catalytic hydrogenation was employed to reduce the double bond and the hydroxyl moiety to give ketone 223. Sodium borohydride mediated reduction of the carbonyl group in 223 gave single diastereomer 224. The THP and methyl ether protecting groups were then removed in a two-step process to give triol 226. The more reactive hydroxyl group on the phenyl ring was then reacted with chloroacetonitrile to furnish nitrile 227. A base mediated hydrolysis of the nitrile provided free acid, treprostinil (228), which was converted to its sodium salt 26 by titration with sodium hydroxide (no yield reported).

Voriconazole (*Vfend*[®])

Voriconazole was launched by Pfizer in both oral and injectable formulations for the treatment of fungal infections in patients intolerant of, or refractory to, other therapy and for the treatment of invasive aspergillosis [92]. It is a triazole antifungal agent whose major mechanism of action is the inhibition of fungal cytochrome P450-mediated 14αlanosterol demethylation [93]. The synthesis [94-96] of voriconazole is an excellent example of process research. As depicted in Scheme 27, 5-fluorouracil (229) was chlorinated in both the 2- and 4- positions using a mixture of phosphorus oxychloride and N,N-dimethylaniline at 95° C to afford 230 in 95% yield. Dichloro pyrimidine 230 was reacted with ethyl magnesium bromide to give dihydropyrimidine adduct 231. Adduct 231 was oxidized prior to quenching using a mixture of iodine and TEA in THF to give 2,4-dichloro-6-ethyl-5-fluoro pyrimidine (232) in 75% yield. Reaction of 232 with two equiv of aqueous NaOH at reflux gave selective displacement of the chloro functionality at 4-position. Acidification of the reaction and extraction with DCM gave 2-chloro-6-ethyl-5-fluoro-4(3H)pyrimidine which was conveniently isolated as its ammonia salt 233. Dechlorination of 233 was achieved using catalytic hydrogenation at 50°C to provide 234 in 80% yield. Alternatively, 4-fluoro-6-ethyl-5-fluoropyrimidine (234) was prepared in a two-pot process in which methyl 3oxopentanoate (235) was fluorinated with fluorine gas to give methyl 2-fluoro-3-oxopentanoate (236) in 80-90% yield [97]. This ester was then cyclized [98] with formamidine acetate in the presence of NaOMe to give 234 in a moderate yield (50-70%). Reaction of 234 with phosphorus oxychloride and TEA afforded 4-chloro-6-methyl-5fluoropyrimidine (237) in 90% yield. Reaction of 237 with NBS in the presence of AIBN initiator provided bromide 238 in 95% yield. A Reformatsky protocol was employed in the condensation of 238 with ketone 239 which was an intermediate in the commercial synthesis of Diflucan [99]. A solution of iodine in THF was added to a slurry of zinc and lead at rt and then a mixture of bromide 238 and ketone 239 were added to the above mixture at 5°C for 30 min. This provided the best diastereomeric selectivity and the ratio of 241 and 240 enantiomeric pair reached approximately 10 to 1. Adduct 241 was de-chlorinated using standard hydrogenation condition (5% w/w Pd on carbon /15 psi hydrogen) to give the racemate of voriconazole. The racemic voriconazole was resolved using (1R)-10-camphorsulfonic acid (242) and crystallization of the required diastereomeric salt provided optically pure voriconazole (28) in 80% yield.

ACKNOWLEDGEMENT

The authors would like to acknowledge the critical evaluation of this review by Dr. M. Y. Chu-Moyer and Dr. S Sakya.

ABBREVIATIONS

ADME Absorption, distribution, metabolism,

excretion

AIBN 2,2'-Azobisisobutyronitrile

BOP Benzotriazole-1-yloxy-

tris(dimethylamino)phosphonium

hexafluorophosphate

BSA Bistrimethyl acetamide

BSTFA Bis(trimethylsilyl)trifluoroacetamide

BSU Bistrimethylsilyl urea

Tetrahydro-1-methyl-3,3-diphenyl-1H,3H-CBS

Pyrrolo[1,2-c][1,3,2]oxazaborole

DCE Dichloroethane **DCM** Dichloromethane

DDH 1,3-Dibromo-5,5-dimethylhydantoin

DDQ 2,3-Dichloro-5,6-dicyano-1,4-benzoquinone

DIBAL-H Diisobutylaluminum hydride

DIPEA Diisopropylethylamine DIPP Diisopropylphosphoryl **DMAP** 4-Dimethylaminopyridine **DMF** *N,N*-Dimethylformamide **DMPU** *N*,*N*'-dimethylpropyleneurea

DMSO Methyl sulfoxide

DPPC Diphenylphosphinic chloride **HOBT** 1-Hydroxybenzotriazole hydrate

I.M. Intramuscularly **IPA** Isopropyl alcohol **IPAC** Isopropyl acetate

Lithium diisopropylamide LDA

NBS N-Bromosuccinimide NCE New chemical entities **NEP** *N*-Ethylpyrrolidinone

1-Methyl-2-pyrrolidinone **NMP NYHA** New York Heart Association

PCC Pyridinium chlorochromate PDC Pyridinium dichromate

TBAF = t-Butyl ammonium fluoride

TBDMS = t-Butyldimethylsilyl

TEA = Triethyl amine

TFA = Trifluoroacetic acid

THF = Tetrahydrofuran
THP = Tetrahydropyran

TIPS = Triisopropyl silyl

TPAP = Tetrapropylammonium perruthenate

TMG = 1,1,3,3-Tetramethylguanidine

p-TSA = para-Toluene sulfonic acid

WSC-HCl = 1-Ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride

REFERENCES

- [1] Graul, A. I. Drug News Perspect **2003**, 16, 22.
- [2] Drug News Perspect. 2002, 15, 57.
- [3] Drug News Perspect. 2002, 15, 113.
- [4] Drug News Perspect. 2002, 15, 457.
- [5] Drug News Perspect. 2003, 16, 48.
- [6] Buti, M.; Esteban, R. Drugs of Today 2003, 39, 127.
- [7] Starrett, J. E.; Mansuri, M. M.; Martin, J. C.; Tortolani, D. R.; Bronson, J. J. EP481214 A1 1992.
- [8] Starrett, J. E.; Tortolani, D. R.; Russell, J.; Hitchcock, M. J. M.; Whiterock, V.; Martin, J. C.; Mansuri, M. M. J. Med. Chem. 1994, 37, 1857.
- [9] Yu, R. H.; Schultze, L. M.; Rohloff, J. C.; Dudzinski, P. W.; Kelly, D. E. Org. Process Res. Dev. 1999, 3, 53.
- [10] Holy, A.; Rosenberg, I.; de Clercq, E. EP253412 B1 1988.
- [11] Salgaller, M. L. Curr. Opin. Oncol. Endoc. Metab. Invest. Drugs 1999, 1, 211.
- [12] Ishizumi, K.; Ohashi, N.; Tanno, N. J. Org. Chem. 1987, 52, 4477.
- [13] Gillard, J. W.; Israel, M. Tetrahedron Lett. 1981, 22, 513.
- [14] Shapiro, D. A.; Renock, S.; Arrington, E.; Chiodo, L. A.; Liu, L.-X.; Sibley, D. R.; Roth, B. L.; Mailman, R. Neuropsychopharmacology 2003, 28, 1400.
- [15] Oshiro, Y.; Sato, S.; Kurahashi, N.; Tanaka, T.; Kikuchi, T.; Tottori, K.; Uwahodo, Y.; Nishi, T. J. Med. Chem. 1998, 41, 658.
- [16] Pollard, C. B.; Wicker, H. T. J. Am. Chem. Soc. 1954, 76, 1853.
- [17] Keating, G. M.; Figgitt, D. P. Drugs 2002, 62, 1899.
- [18] Zeitlin, A. L.; Stirling, D. I. US5733756 A 1998.
- [19] Prashad, M.; Har, D. *US6100401 A* **2000**.
- [20] Prashad, M.; Hu, B. *US6162919 A* **2000**.
- [21] Thai, D. L.; Sapko, M. T.; Reiter, C. T.; Bierer, D. E.; Perel, J. M. J. Med. Chem. 1998, 41, 591.
- [22] Graul, A.; Silvestre, J.; Castañer, J. Drugs Future 1999, 24, 246.
- [23] Davis, R.; Millar, A.; Sterbenz, J. T. WO0246207 A2 2002.
- [24] Odenholt, I. Exp. Opin. Invest. Drugs **2001**, 10, 1157.
- [25] Berks, A. H. Tetrahedron 1996, 52, 331.
- [26] Carbapenem enolphosphate 71 is commercial available from Takasago, Kaneka and Nisso companies.
- [27] Brands, K. M. J.; Jobson, R. B.; Conrad, K. M.; Williams, J. M.; Pipik, B.; Cameron, M.; Davies, A. J.; Houghton, P. G.; Ashwood, M. S.; Cottrell, I. F.; Reamer, R. A.; Kennedy, D. J.; Dolling, U.-H.; Reider, P. J. J. Org. Chem. 2002, 67, 4771.
- [28] Williams, J. M.; Skerlj, R. WO02057266 A1 2002.
- [29] Burke, W. J. Exp. Opin. Invest. Drugs 2002, 11, 1477.
- [30] Boegesoe, K. P.; Perregaard, J. US4943590 1990.
 [31] Boegesoe, K. P.; Toft, A. S. US4136193 1979.
- [31] Boegesoe, K. P.; 10ft, A. S. US4130193 1979. [32] Ahmadian, H.; Petersen, H. WO03051861 2003.
- [33] Boegesoe, K. P. *US4650884* **1987**.
- [34] Cochrane, D. J.; Jarvis, B.; Keating, G. M. Drugs 2002, 62, 2637.
- [35] Davies, I. W.; Marcoux, J.-F.; Corley, E. G.; Journet, M.; Cai, D.-W.; Palucki, M.; Wu, J.; Larsen, R. D.; Rossen, K.; Pye, P. J.; DiMichele, L.; Dormer, P.; Reider, P. J. J. Org. Chem. 2000, 65, 8415.
- [36] Harris, M.; Davis, W.; Brown, W. V. Drugs of Today 2003, 39, 229.

- [37] Wu, G.; Wong, Y.; Chen, X.; Ding, Z. J. Org. Chem. 1999, 64, 3714
- [38] Thiruvengadam, T. K.; Fu, X.; Tann, C. H.; McAllister, T. L.; Chiu, J. S.; Colon, C. US6207822 2001.
- [39] Fu, X.; McAllister, T. L.; Thiruvengadam, T. K.; Tann, C. -H.; Su, D. Tetrahedron Lett. 2003, 44, 801.
- [40] Cheng, J. W. M. Clin. Therap. 2002, 24, 1757.
- [41] Petitou, M.; Duchaussoy, P.; Jaurand, G.; Gourvenec, F.; Lederman, I.; Strassel, J. M.; Barzu, T.; Crepon, B.; Herault, J., P.; Lormeau, J. C.; Bernat, A.; Herbert, J. M. J. Med. Chem. 1997, 40, 1600.
- [42] van Boeckel, C. A. A.; Petitou, M. Angew. Chem. Int. Ed. 1993, 32, 1671.
- [43] Petitou, M.; Duchaussoy, P.; Lederman, I.; Choay, J.; Jacquinet, J. C.; Sinay, P.; Torri, G. Carbohydrate Res. 1987, 167, 67.
- [44] Petitou, M.; Jaurand, G.; Derrien, M.; Duchaussoy, P.; Choay, J. Bioorg. Med. Chem. Lett. 1991, 1, 95.
- [45] Easthope, S. E.; Goa, K. L. CNS Drugs 2001, 15, 969.
- [46] Brackenridge, I.; Spray, C.; McIntyre, S.; Knight, J.; Hartley, D. WO9954302 A1 1999.
- [47] Howell, A.; Robertson, J. F. R.; Albano, J. Q.; Aschermannova, A.; Mauriac, L.; Kleeberg, U. R.; Vergote, I.; Erikstein, B.; Webster, A.; Morris, C. J. Clin. Oncol. 2002, 20, 3396.
- [48] Warren, K. E. H.; Kane, A. M. L. WO03031399 A1 2003.
- [49] Bowler, J.; Lilley, T. J.; Pittam, J. D.; Wakeling, A. E. *Steroids* **1989**, 54, 71.
- [50] Stevenson, R.; Kerr, F. W.; Lane, A. R.; Brazier, E. J.; Hogan, P. J.; Laffan, D. D. P. WO0232922 A1 2002.
- [51] Culy, C. R.; Faulds, D. Drugs 2002, 62, 2237.
- [52] Barker, A. EP566226 B1 1995.
- [53] Gibson, K. EP823900 B1 2000.
- [54] Junichi, O.; Takashi, O.; Kouichiro, M. Can. J. Anaesthesia 2003, 50, 753.
- [55] Iguchi, S.; Kawamura, M.; Miyamoto, T. EP397031 A1 1990.
- [56] Fromtling, R. A. Drugs of Today 2002, 38, 245.
- [57] Tomishima, M.; Ohki, H.; Yanada, A.; Takasugi, H.; Maki, K.; Tawara, S.; Tanaka, H. J. Antibiotics 1999, 52, 674.
- [58] Adami, S.; Gatti, D.; Colapietro, F.; Fracassi, E.; Braga, V.; Rossini, M.; Tato, L. J. Bone Mineral Res. 2003, 18, 126.
- [59] Guainai-Ricci, G.; Rosini, S. EP494844 B1 1992.
- [60] Holme, E.; Lindstedt, S. J. Inh. Metab. Disease 1998, 21, 507.
- [61] Bay, E. US4774360 A 1988.
- [62] Brousil, J. A.; Burke, J. M. Clin. Therap. 2003, 25, 1041.
- [63] Yanagisawa, H.; Fujimoto, K.; Amemiya, Y.; Shimoji, Y.; Kanazaki, T.; Koike, H.; Sada, T. US5616599 A 1997.
- [64] Malan, T. P., Jr.; Marsh, G.; Hakki, S. I.; Grossman, E.; Traylor, L.; Hubbard, R. C. Anesthesiology 2003, 98, 950.
- [65] Letendre, L. J.; Kunda, S. A.; Gallagher, D. J.; Seaney, L. M. WO 03029230 A1 2003.
- [66] Talley, J. J.; Bertenshaw, S. R.; Brown, D. L.; Carter, J. S.; Graneto, M. J.; Kellogg, M. S.; Koboldt, C. M.; Yuan, J. H.; Zhang, Y. Y.; Seibert, K. J. Med. Chem. 2000, 43, 1661.
- [67] Nomura, N.; Mitsuyama, J.; Furuta, Y.; Yamada, H.; Nakata, M.; Fukuda, T.; Yamada, H.; Takahata, M.; Minami, S. Jpn. J. Antib. 2002, 55, 412.
- [68] Todo, Y.; Takagi, H.; Iino, F.; Hayashi, K.; Takata, M.; Kuroda, H.; Momonoi, K.; Narita, H. Chem. Pharm. Bull. 1994, 42, 2629.
- [69] Graham-Brown, R.; Grassberger, M. Int. J. Clin. Practice 2003, 57,
- [70] Fleissner, G.; Hacker, H.; Kusters, E.; Penn, G. W00190110 A1
- [71] Dosenbach, C.; Grassberger, M.; Hartmann, O.; Horvath, A.; Mutz, J.-P.; Penn, G.; Pfeffer, S.; Wieckhusen, D. WO9901458 A1 1999.
- [72] Baumann, K.; Emmer, G. EP427680 B1 1991.
- [73] Bochis, R. J.; Wyvratt, Jr., M. J. EP480623 A1 1992.
- [74] Okuhara, M.; Tanaka, H.; Goto, T. EP0184162 B1 1986.
- [75] Jones, T. K.; Mills, S. G.; Reamer, R. A.; Askin, D.; Desmond, R.; Volante, R. P.; Shinlai, I. J. Am. Chem. Soc. 1989, 111, 1157.
- [76] Barrett, J. F. Curr. Opin. Anti-Infect. Invest. Drugs 1999, 1, 453.
- [77] Segawa, J.; Kitano, M.; Kazuno, K.; Matsuoka, M.; Shirahase, I.; Ozaki, M.; Matsuda, M.; Tomii, Y.; Kise, M. J. Med. Chem. 1992, 35, 4727
- [78] Matsuoka, M.; Segawa, J.; Makita, Y.; Ohmachi, S.; Kashima, T.; Nakamura, K.; Hattori, M.; Kitano, M.; Kise, M. J. J. Heterocycl. Chem. 1997, 34, 1773.
- [79] Schuster, H. Cardiology 2003, 99, 126.
- [80] Watanabe, M.; Koike, H.; Ishiba, T.; Okada, T.; Seo, S.; Hirai, K. Bioorg. & Med. Chem. 1997, 5, 437.

- [81] Konoike, T.; Araki, Y. J. Org. Chem. 1994, 59, 7849.
- [82] Pradella, L. IDrugs 2000, 3, 208.
- [83] Imaki, K.; Wakatsuka, H. EP539223 A1 1993.
- [84] Imaki, K.; Okada, T.; Nakayama, Y.; Nagao, Y.; Kobayashi, K.; Sakai, Y.; Mohri, T.; Amino, T.; Nakai, H.; Kawamura, M. *Bioorg. Med. Chem.* 1996, 4, 2115.
- [85] Panning, C. A.; DeBisschop, M. Pharmacotherapy 2003, 23, 183.
- [86] Banholzer, R.; Bauer, R.; Reichl, R. EP418716 A1 1991.
- [87] Banholzer, R.; Bauer, R.; Reichl, R. US5610163 A 1997.
- [88] Banholzer, R.; Graulich, M.; Luettke, S.; Mathes, A.; Meissner, H.; Specht, P.; Broeder, W. US20020133010 A1 2002.
- [89] Horn, E. M.; Barst, R. J. Exp. Opin. Invest. Drugs 2002, 11, 1615.
- [90] Moriarty, R. M.; Penmasta, R.; Guo, L.; Rao, M. S.; Staszewski, J. P. US 6441245 B1 2002.
- [91] Moriarty, R. M.; Penmasta, R.; Guo, L.; Rao, M. S.; Staszewski, J. P. WO9921830 1999.

- [92] Gunderson, S. M.; Jain, R.; Danziger, L. H. J. Pharm. Tech. 2003, 19 97
- [93] Van Epps, H. L.; Feldmesser, M.; Pamer, E. G. Antimicrob. Agents Chemotherapy 2003, 47, 1818.
- [94] Bartroli, J.; Turmo, E.; Algueró, M.; Boncompte, E.; Vericat, M. L.; Conte, L.; Ramis, J.; Merlos, M.; García-Rafanell, J.; Forn, J. J. Med. Chem. 1998, 41, 1869.
- [95] Butters, M.; Ebbs, J.; Green, S. P.; MacRae, J.; Morland, M. C.; Murtiashaw, C. W.; Pettman, A. J. Org. Process Res. Dev. 2001, 5, 28.
- [96] Butters, M.; Harrison, J. A.; Pettman, A. J. WO9706160 A1 1997.
- [97] Nukui, K.; Fukami, S.; Kawada, K. WO9735824 A1 1997.
- [98] Butters, M. J. Heterocycl. Chem. 1992, 1369.
- [99] Dickinson, R. P.; Bell, A. S.; Hitchcock, C. A.; Narayanaswami, S.; Ray, S. J.; Richardson, K.; Troke, P. F. Bioorg. Med. Chem. Lett. 1996, 6, 2031.

Copyright of Mini Reviews in Medicinal Chemistry is the property of Bentham Science Publishers Ltd. and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.

Copyright of Mini Reviews in Medicinal Chemistry is the property of Bentham Science Publishers Ltd. and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.